

chemically induced skin tumors, the aromatic analog I was similarly inactive as retinoic acid when tested against transplantable tumors like Ehrlich carcinoma, solid form, Ehrlich ascites tumor, Crocker sarcoma S 180 or leukemia L 1210 (BOLLAG⁹). Beside the direct antipapilloma effect, a metaplasia-preventing effect of I on the vaginal epithelium of the rat could be demonstrated in the modified colpokerososis test of BOGUTH et al.¹⁰ With equimolar doses I showed 79% of the activity of retinoic acid (WEISER¹¹). It would be very interesting to know whether the metaplasia or dysplasia preventing effect could be demonstrated also in other systems like organ cultures of tracheal epithelium¹² or prostate epithelium¹³.

We may conclude that I, a retinoic acid analog, has a marked therapeutic effect on chemically induced benign and malignant epithelial tumors. With respect to the papilloma-regressing effect, I is superior to retinoic acid, as reflected in a 10 times better therapeutic ratio. This investigation has proved that the antitumor effect is not

strictly linked with the development of hypervitaminosis A and that a dissociation between these two properties leads to a broader therapeutic margin. In preliminary studies it could be demonstrated that the aromatic analog I exerts also the same superiority over retinoic acid when given prophylactically. I delayed markedly the induction of premalignant as well as malignant epithelial skin lesions⁹. Clinical trials are undertaken on the therapy of precancerous conditions in man, with the goal to reach thereby a prophylaxis of malignant epithelial tumors.

Zusammenfassung. Retinsäure hat einen prophylaktischen und therapeutischen Effekt auf chemisch induzierte benigne und maligne epitheliale Tumoren. Die Wirkungen werden durch das Auftreten der sogenannten Hypervitaminose A-Symptome limitiert. Es werden die biologischen Eigenschaften eines aromatischen Retinsäure-Analogen (Figur) beschrieben, bei dem das Verhältnis zwischen den Dosen, die eine Tumorerregung bewirken, und denen, die eine Hypervitaminose A erzeugen, 10mal günstiger ist als bei Retinsäure. Papillomregression und Hypervitaminose A sind nicht eng miteinander gekoppelt. Eine Dissoziation dieser biologischen Eigenschaften führt zu Substanzen mit besserem therapeutischem Quotient.

W. BOLLAG

⁹ W. BOLLAG, unpublished.

¹⁰ W. BOGUTH, V. HORN, M. K. SOLLIMAN and H. WEISER, *Int. Z. Vitaminforsch.* 37, 6 (1960).

¹¹ H. WEISER, personal communication.

¹² G. H. CLAMON, M. B. SPORN, J. M. SMITH and U. SAFFIOTTI, *Nature, Lond.* 250, 64 (1974).

¹³ I. LASNITZKI and DE WITT S. GOODMAN, *Cancer Res.* 34, 1564 (1974).

*Department of Experimental Medicine,
F. Hoffmann-La Roche & Co. Ltd.,
CH-4002 Basel (Switzerland), 15 May 1974.*

On the Modification of the Divalent Cation-Binding by Phospholipids in Tumours

Phospholipids are the major structural components of the cell membrane and their combination with calcium and magnesium play a role of vital importance in many physiological phenomena, such as cell secretion¹, active transport², and cell permeability³. The isolation of phospholipid-calcium complexes from experimental tumours has been already reported⁴.

A comparative study on the concentration of calcium and magnesium complexes with phospholipid in hepatoma, liver of hepatoma-bearing animals and liver from normal animals was carried out in the present experimental work.

8-day-old Novikoff hepatoma (ascitic or solid form) and 30-day-old hepatoma BW7756 bearing animals were used in this investigation. Groups of normal Holzman rats and C57L/J mice of the same sex and weight were used as control groups. In order to isolate the phospho-

¹ W. V. DOUGLAS and R. P. RUBIN, *J. Physiol., Lond.* 159, 40 (1961).

² G. GARDOS, *Acta physiol. hung.* 18, 265 (1961).

³ L. V. HEILBRUNN, in *An Outline of General Physiology* (W. B. Saunders & Co., Philadelphia 1952).

⁴ L. J. ANGHILERI, *Experientia* 28, 1086 (1972).

Calcium and magnesium complexed by the phospholipids of neoplastic and normal tissues

| | Total lipid phosphorus ($\mu\text{g P/g tissue}$) | Complexed phospholipid (% of total lipid P) | Complexed calcium ($\mu\text{g Ca/g tissue}$) | Complexed magnesium ($\mu\text{g Mg/g tissue}$) |
|--------------------------------|--|--|--|--|
| Novikoff hepatoma ^b | | | | |
| Solid form | 114 (91–156) * | 9.2 (5.8–14.0) * | 3.9 (2.2–6.3) * | 1.1 (0.7–1.6) * |
| Ascites form | 201 (190–223) | 8.3 (6.5–11.0) | 1.7 (1.0–3.1) | 2.9 (1.8–4.2) |
| Liver solid tumor bearing | 340 (232–424) | 6.3 (2.7–12.9) | 3.5 (2.7–4.2) | 5.8 (2.8–7.3) |
| Liver ascites bearing | 340 (241–451) | 13.0 (10.8–16.6) | 4.5 (4.1–5.2) | 8.1 (7.2–9.3) |
| Liver normal animals | 436 (386–500) | 12.7 (6.4–18.8) | 4.2 (2.8–5.5) | 10.1 (7.9–11.4) |
| Hepatoma BW7756 ^c | | | | |
| Solid tumor | 23 (15–28) | 7.4 (5.2–9.0) | 2.5 (1.9–3.0) | 1.5 (1.2–1.6) |
| Liver tumor bearing | 66 (51–71) | 9.6 (7.2–11.1) | 3.3 (3.0–3.9) | 4.8 (4.3–5.7) |
| Liver normal animals | 95 (93–96) | 7.3 (6.4–8.9) | 2.1 (1.9–2.4) | 3.6 (3.4–3.8) |

*, Range; ^b, from 20 animals; ^c, from 45 animals.

lipid complexes, the tumour and liver were processed as described elsewhere⁴. After mineralization of the fractions obtained with HClO_4 plus HNO_3 , phosphorus, calcium and magnesium were measured in aliquots of the mineralized material. Phosphorus was assayed colorimetrically using aminonaphtolsulfonic acid reagent⁵. Calcium and magnesium were determined with an Atomic Absorption Spectrometer. The results shown in the Table are the average and the range values corresponding to each type of tumour.

The observation that tumor tissue contains a much lower content of phospholipid than liver is in agreement with that of other authors^{6,7}. The most striking difference between hepatoma and normal liver is a much lower amount of magnesium complexed by the phospholipids, while the complexed calcium does not show significant variations. Since it has been demonstrated that the binding of calcium and magnesium by the membranes exhibits a saturation type relationship characteristic of adsorption to binding sites and describable by the law of mass action⁸, this difference may be due to a considerably higher calcium concentration at the cell membrane level. On the other hand, considering that tumor has shown a slightly higher proportion of cephalin than liver⁹, and that in biphasic systems the acidic phospholipids have shown a far greater affinity for Ca^{2+} than for Mg^{2+} ¹⁰, it is possible that this small difference in the phospholipid composition may also contribute to the magnesium-binding behavior observed.

These findings raise the question whether this change in the divalent cation distribution may or may not be implicated in determining the tumor cell membrane characteristics. In relation to this, it is also very interesting to note that, with less than half of the complexed

phospholipid amount, tumors bind almost as much calcium as liver does. This is an indication of a drastic change in the phospholipid-calcium relationship.

Zusammenfassung. Phospholipid-Kalzium- und Phospholipid-Magnesium-Komplexe wurden aus Hepatom und aus normaler Leber isoliert. Obwohl das Hepatom nur die Hälfte der in der Leber vorkommenden komplexen Phospholipide aufweist, ist die Kalziumbindung bei Hepatom und Leber gleich. Die Magnesiumbindung ist dagegen erheblich niedriger. Diese Unterschiede weisen auf eine mögliche Bedeutung der zweiwertigen Kationenbindung im Verhalten der Tumorzellmembran hin.

L. J. ANGHILERI¹¹

*Innere Klinik und Poliklinik (Tumorforschung),
Klinikum der GHS Essen, Hufelandstrasse 55,
D-4300 Essen (Federal Republic of Germany),
28 May 1974.*

⁵ G. F. SHINOWARA, L. M. JONES and H. L. REINHART, *J. biol. Chem.* **142**, 921 (1942).

⁶ J. H. VEERKAMP, I. MULDER and I. VAN DEENEN, *Z. Krebsforsch.* **64**, 137 (1961).

⁷ P. H. FIGARD and D. M. GREENBERG, *Cancer Res.* **22**, 361 (1962).

⁸ A. P. CARVALHO, H. SANUI and N. PACE, *J. cell. comp. Physiol.* **62**, 311 (1963).

⁹ R. M. JOHNSON and P. H. DUTCH, *Archs Biochem. Biophys.* **40**, 239 (1952).

¹⁰ H. HAUSER and R. M. C. DAWSON, *Europ. J. Biochem.* **7**, 61 (1967).

¹¹ This work has been supported by the Ministerium für Wissenschaft und Forschung des Landes Nordrhein-Westfalen.

On the Quarternary Structure of *Carcinus moenas* (Arthropoda) Hemocyanin^{1,2}

It is well enough established that the so-called minimum functional subunit of the hemocyanin (Hc) of several Arthropoda species (unlike Mollusca³) can be isolated without covalent bond cleavage^{4,5}. This component includes one active site with 2 copper ions, weighs around 75,000 d and has been reported to be rather heterogeneous⁶⁻⁹. Three different hypotheses about its constitution have been proposed following the studies on minimum subunits regardless of their functionality: 1. PICKETT, RIGGS and LARIMER¹⁰ have reported that succinylated *Homarus americanus* Hc displays by sedimentation analysis a subunit of about 37,500 d, which probably consists of only one polypeptide chain; 2. according to another model¹¹, suggested in order to explain the low and 'continuously varying' sedimentation coefficient of *Cancer magister* Hc in 6 M guanidine hydrochloride solution, the minimum functional subunit is composed of 3 polypeptide chains of 25,000 d. Such a model was made probable by recent data obtained in our laboratory on *Carcinus moenas* Hc by polyacrylamide gel electrophoresis and gel permeation chromatography in 0.1% SDS solutions at pH 7.0-9.3^{12,13}; 3. the results of LOEHR and MASON⁸, confirmed by CARPENTER and VAN HOLDE⁹, seem to prove that the subunits around 80,000 d of *Cancer magister* Hc consist of single polypeptide chains. These results have been obtained by polyacrylamide gel electrophoresis in sodium dodecyl sulfate (SDS) solutions at pH 8.0-9.0.

We have separated *Carcinus moenas* Hc polypeptide chains using gel permeation chromatography on Sephadex

G-200 in 1% SDS solution^{14,15}. Hc and apoHc (apo-hemocyanin) were prepared according to GHIRETTI

¹ A preliminary report on this work was communicated at the 10th National Meeting of the Italian Society of Biophysics and Molecular Biology (Padua, 28/9/1973).

² This work was supported by CNR grants.

³ R. LONTIE, M. DE LEY, H. ROBBERECHT and R. WITTERS, *Nature, Lond.* **242**, 180 (1973).

⁴ K. E. VAN HOLDE and E. F. J. VAN BRUGGEN, in *Biol. Macromolecules Series* (Eds. N. TIMASHEFF and G. D. FASMAN; Dekker, New York 1971), vol. 5, p. 1.

⁵ F. GHIRETTI, A. GHIRETTI MAGALDI and B. SALVATO, in *Comparative Physiology* (Eds. L. BOLIS, K. SCHMIDT-NIELSEN and S. H. P. MADDRELL; North-Holland Publishing Co. Amsterdam 1973), p. 509.

⁶ L. DI GIAMBERARDINO, *Archs Biochem. Biophys.* **118**, 273 (1967).

⁷ P. BUSSELEN, *Archs Biochem. Biophys.* **137**, 415 (1970).

⁸ J. S. LOEHR and H. S. MASON, *Biochem. biophys. Res. Commun.* **51**, 741 (1973).

⁹ D. E. CARPENTER and K. E. VAN HOLDE, *Biochemistry* **12**, 2231 (1973).

¹⁰ S. M. PICKETT, N. F. RIGGS and J. L. LARIMER, *Science* **151**, 1005 (1966).

¹¹ H. D. ELLERTON, D. E. CARPENTER and K. E. VAN HOLDE, *Biochemistry* **9**, 2225 (1970).

¹² B. SALVATO, S. SARTORE, G. ZACCARIA and A. GHIRETTI MAGALDI, *Boll. Soc. ital. Biol. sper.* **47**, 777 (1971).

¹³ B. SALVATO, S. SARTORE, M. RIZZOTTI and A. GHIRETTI MAGALDI, *FEBS Lett.* **22**, 5 (1972).

¹⁴ W. W. FISH, J. A. REYNOLDS and C. TANFORD, *J. biol. Chem.* **245**, 5166 (1970).

¹⁵ K. WEBER and D. J. KUTER, *J. biol. Chem.* **246**, 4504 (1971).